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► To cite this version:

Wenyu Zhang, Weixin Zhang, Daokuo Ge, Hongxin Cao, Yan Liu, et al.. Biomass-Based Leaf Curvilinear Model for Rapeseed (*Brassica napus* L.). 9th International Conference on Computer and Computing Technologies in Agriculture (CCTA), Sep 2015, Beijing, China. pp.459-472, 10.1007/978-3-319-48357-3_44 . hal-01557840

HAL Id: hal-01557840

<https://inria.hal.science/hal-01557840>

Submitted on 6 Jul 2017

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Biomass-Based Leaf Curvilinear Model for Rapeseed (*Brassica napus* L.)

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Abstract: Leaf is one of the most important photosynthetic organs of rapeseed (*Brassica napus* L.). To quantify relationships between the leaf curve and the corresponding leaf biomass for rapeseed on main stem, this paper presents a biomass-based leaf curvilinear model for rapeseed. Various model variables, including leaf length, bowstring length, tangential angle, and bowstring angle, were parameterized based on data derived from the field experiments with varieties, fertilizer, and transplanting densities during 2011 to 2012, and 2012 to 2013 growing seasons. And then we analysed the biological significance of curvilinear equation for straight leaves, constructed the straight leaf probabilistic model on main stem, quantified the relationship between leaf curvature and the corresponding leaf biomass, and constructed the leaf curvilinear model based on the assumption and verification of the curvilinear equation form for curving leaf. The probability of straight leaf can be quantified with piecewise function according to the different trend in the normalized leaf ranks ((0, 0.4], and (0.4, 1]). The leaf curvature decreased with the increasing of leaf biomass, and can be described with reciprocal function. The curve of straight leaf and the curving leaf can be simulated by linear equation and the quadratic function, respectively. Our models were validated with the independent dataset from the field experiment, and the results indicated that the model could effectively predict the straight leaf probability and leaf curvature, which would be useful for linking the rapeseed growth model with the rapeseed morphological model, and set the stage for the development of functional-structural rapeseed models.

Keywords: rapeseed (*Brassica napus* L.), biomass, leaf curve, functional-structural plant models (FSPMs)

1 Introduction

Rapeseed is the world's important oil crops [1] with harvest area of 25.3 to 30.9 million ha and total yield of 46.5~72.5 million tons during 2004~2013 [2]. At the same time, it is the main oilseed crop in China [3], whose harvest area is about 5.6~7.5 million ha, and the total yield is about 10.6~14.4 million tons [2] in general. Also, it is one of the main raw material of biodiesel [4]. Therefore, it is very important for ensure food and ecological security that promote the development of rapeseed production.

Light distribution characteristics in crop canopies directly affect the light energy utilization efficiency for photosynthesis, dry matter accumulation, and yield formation. Almost all the growth models predicted the crop canopy light distribution through the Beer's law [5-8], in that the two key factors for light distribution simulation process, the extinction coefficient and the layered leaf area index, are closely

related with the leaf curving characteristics [9]. Therefore, quantitatively modeling of the leaf curve could provide a mechanistic way for precisely simulating the crop canopy structure, light distribution, and photosynthesis, and lay a foundation for the predicting of light energy utilization efficiency and yield formation.

At present, there are many studies on leaf curve modeling. In the study of mathematical characterization of maize canopies [10], leaf curve was described as a general quadratic equation expressed by the initial leaf angle, the coordinates of the leaf tip and the leaf's maximum height. The general quadratic equation was also used to simulate the leaf curve of maize [11-13], rice [14], and other crops by many researchers. Leaf curve was also fitted into a quadratic function for rice [15, 16] and winter wheat [17], or a Gaussian function for spring barley [18] and rice [19]. Furthermore, Espana et al. [20] decomposed leaf curvature into two parts, the ascending part was described as a parabolic curve, and the descending part, when existing, was characterized by a portion of an ellipse, and then applied the leaf curvature model to maize canopy 3D architecture and reflectance simulation. Watanabe et al. [21] found that leaf curves could be fitted using Hermite functions through analyzing three angles related to the basal, mid, and tip of leaf. Shi et al. [22] characterized the rice leaf curve by a second order differential equation, including the synthetic effect of leaf blade length, width, specific leaf weight, initial leaf angle, and the deformation coefficient on leaf space shape, using force analyzing on rice leaf. Zheng et al. [23] obtain leaf midrib coordinate points by cubic spatial B-spline interpolation, and characterized the leaf curves as the connecting line of these points.

It is difficult to measure leaf curve because of the intricate leaf shape for rapeseed. Therefore, the objectives of this research were to develop straight leaf probabilistic model, straight leaf curvilinear model, and biomass-based leaf curvilinear model by linking leaf morphological parameters with the corresponding leaf biomass, to validate the hypothesis that the curvilinear function for curving leaves could be fitted as quadratic function, and to provide a reference for linking morphological parameters with corresponding organ biomass, and for the establishment of the FSPMs.

2 Materials and Methods

2.1 Materials

We used 2 rapeseed cultivars, they are: "Ningyou 18" (V1, conventional), and "Ningza 19" (V2, hybrid), bred by Institute of Economic Crops Research, Jiangsu Academy of Agricultural Sciences.

2.2 Methods

2.2.1 Experimental Conditions and Design

In order to determine the parameters and verify the models, three experiments were conducted involving different varieties, transplanting densities, and fertilizer during the 2011-2012, and 2012-2013 growing seasons at the experimental farm of our Academy (32.03 °N, 118.87 °E). The soil type is a hydric anthrosol (organic carbon, 31.4 g kg⁻¹; total nitrogen, 2.03 g kg⁻¹; available phosphorus, 20.3 mg kg⁻¹; available potassium, 139.0 mg kg⁻¹; and pH 7.31).

Exp. 1, variety and the fertilizer experiment (2011-2012): The Experiment was deployed in split block design with three replications. Two fertilizer levels (N0 = no fertilizer; N2 = 180 kg ha⁻¹) were the whole-plot treatments while two cultivars (V1 and V2) constituted the sub-plots. The plots arranged

random with 0.4 m row spacing, 0.17-0.20 m plant spacing in 7.0×5.7 m area. Fertilizer contained 12kgP₂O₅ha⁻¹, 18kgK₂Oha⁻¹, and 15 kgboron ha⁻¹.

Exp. 2, variety experiment (2012-2013): The experiment was deployed in randomized complete block design with 2 varieties (V1 and V2) and 3 replications. Nitrogen fertilizer included 90kg N ha⁻¹ (N1), and the transplanting density was 1.2×10⁵ plantha⁻¹ (D2).

Exp. 3, variety, fertilizer and transplanting density experiment (2012-2013): The Experiment was deployed in split block design with three replications. Three fertilizer levels (N0 = no fertilizer; N1 = 90kg ha⁻¹; N2 = 180kg ha⁻¹) were the whole-plot treatments while variety (V1) and three transplanting densities (D1= 6×10⁴ plantha⁻¹; D2 = 1.2×10⁵ plantha⁻¹; D3= 1.8×10⁵ plantha⁻¹) constituted the sub-plots. The plots of Exp. 2 and Exp. 3 arranged random with 0.42 m row spacing in 3.99 by 3.5 m area, and the plant spacing was calculated by row spacing and transplanting density. Fertilizer contained 90kgP₂O₅ha⁻¹ and 90kgK₂Oha⁻¹ for N1 plots, and 180kgP₂O₅ha⁻¹ and 180 kgK₂Oha⁻¹ for N2 plots, and 15 kgboron ha⁻¹ was used as foliage spray for both N1 and N2 plots after bolting.

2.2.2 Measurements

The leaf rank on the main stem of 50 randomly selected seedlings for each plot were marked using a red number stamp before transplanting. Leaf morphological parameters including leaf length (the distance between leaf basal and leaf tip in the straight state, including the leaf blade and petiole, if it exists. *LL*, for short), leaf tangential angle (the angle between the tangential direction of leaf basal and the main stem. *TA*, for short), leaf bowstring angle (the angle between the straight line from leaf basal to leaf tip in natural state and the main stem. *BA*, for short), and leaf bowstring length (the distance between leaf basal and leaf tip in natural state. *LBL*, for short) were measured using a straight edge and protractor directly (Fig. 1).

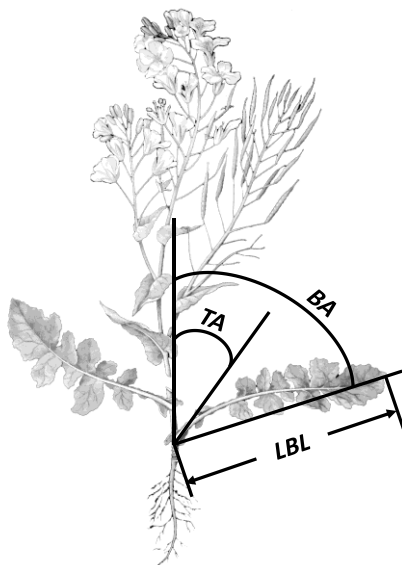


Fig. 1. Diagram of *TA*, *BA* and *LBL*

2.2.3 Data Analysis

We wrote a C# Program to solve the approximate solution of leaf curve equation parameters through a step of 10⁻⁵ cm. Leaf rank data were normalized to (0, 1] interval, in order to eliminate the apparent differences between treatments and replications. The data from Exp. 1 was used for model development, data from Exp. 2, and Exp. 3 were used for validation.

2.2.4 Model Validation

We validated the models developed in this paper by calculating the correlation (r), the root mean square error ($RMSE$), the average absolute difference (d_a), and the ratio of d_a to the average observation (d_{ap})[24], and 1:1 line of simulated and observed properties. Some statistical indices were defined as follows:

$$RMSE = \sqrt{\frac{\sum_{i=1}^n (O_i - S_i)^2}{n}}$$

$$d_a = \frac{1}{n} \sum_{i=1}^n (O_i - S_i)$$

$$d_{ap}(\%) = |d_a|/\bar{O} \times 100$$

where i is sample number, n is total number of measurements, $n-1=n$ when $n \geq 30$, S_i is simulated value, and O_i is observed value.

3 Results

3.1 Model Description

3.1.1 Probability and Curvilinear Model for Straight Leaves

In order to represent the extension state of leaf better, we set the leaf in the Cartesian coordinate system with the leaf basal point as the origin and growth direction of main stem as y-axis, regardless of leaf distorting. According to observations in the Exp. 1, some rapeseed leaves could be considered as straight leaves with small difference between leaf length and leaf bowstring length, as well as leaf tangential angle and leaf bowstring angle. Therefore, we treated the leaves as straight leaves if difference between the leaf length and the leaf bowstring length was less than 1 cm or difference between the leaf tangential angle and the leaf bowstring angle is less than 10° . The curvilinear equation ($f(x)$) of straight leaves could be expressed as a linear function which passes through the origin and with the cotangent value of the leaf tangential angle as the slope, and the function could be described by Eq. (1).

$$f(x) = \cot(TA) \cdot x \quad (1)$$

where TA can be simulated by our previous model[25].

The data in the Exp. 1 showed that changes in the probability of different treatments for straight leaves by the normalized leaf rank was close to quadratic curve with significant r ($r=0.725$, $P<0.01$, $n=16$, $r_{(14, 0.01)}=0.623$, Table 1) in the interval (0, 0.4] (Fig. 2a), and logarithmic curve with significant r ($r=0.925$, $P<0.001$, $n=33$, $r_{(31, 0.001)}=0.547$, Table 1) in the interval (0.4, 1] (Fig. 2b). So that the probability of straight leaves by the normalized leaf ranks (P_{LS}) could be expressed as a piecewise function as Eq. 2.

$$P_{LS} = \begin{cases} A_1 \cdot NLRs^2 + B_1 \cdot NLRs + C_1 & NLRs \in (0, 0.4] \\ A_2 \cdot \ln(NLRs) + B_2 & NLRs \in (0.4, 1] \end{cases} \quad (2)$$

where $NLRs$ is normalized leaf rank; A_1, A_2, B_1, B_2 , and C_1 are model parameters whose values and testing data shown in Table 1.

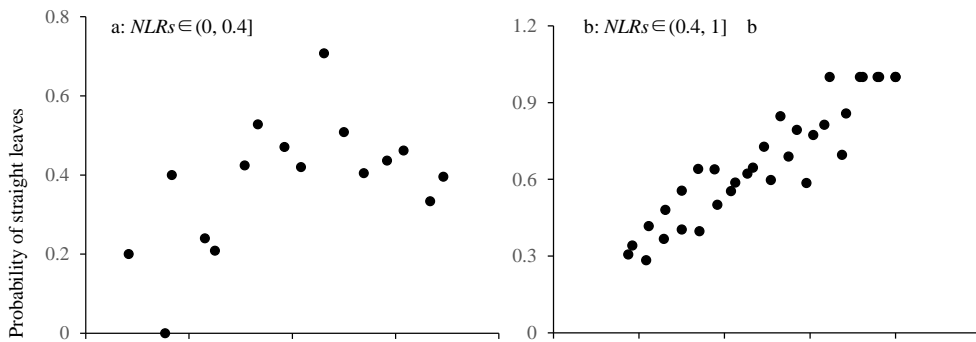


Fig. 2. Changes in the probability of straight leaves with the normalized leaf ranks

Table 1Significance test of the straight leaf probabilistic model and its parameters

Interval of normalized leaf ranks	Function	<i>n</i>	<i>r</i>	Sig. for <i>r</i>	Sig. for <i>F</i>	Parameter symbolic	Unstandardize d coefficients	t
(0, 0.4]	$A_1 \cdot NLRs^2$	16	0.725 ^{**}	$r_{(14, 0.01)}=0.623$	0.008	A_1	-10.519	-2.709 [*]
	$+ B_1 \cdot NLRs$					B_1	1.572	3.199 ^{**}
	$+ C_1$					C_1	0.139	-0.800
(0.4, 1]	$A_2 \cdot \ln(NLRs)$	33	0.925 ^{**}	$r_{(31, 0.001)}=0.547$	0.000	A_2	0.716	13.575 ^{***}
	$+ B_2$					B_2	0.965	36.223 ^{***}

***, **, and * denote significance at $P<0.001$, $P<0.01$, and $P<0.05$, respectively. The same as below.

3.1.2 CurvilinearModel for CurvingLeaves

3.1.2.1AssumedFunctions and There Biological Significance for Leaf Curve

According to the leaves curving characteristicsobserved in experiments,and the research on curvilinear models for other crops, we supposedthat curvilinear function of rapeseedleaf ($f(x)$) could be fitted by functions such asquadratic function(ax^2+bx),quartic function($ax^4+bx^3+cx^2+dx$), sine function($asin(bx)$), Hoerl-like function(axb^x),($a+bx^c$)/($d+x^c$),and $(a+bx)/(1+cx+dx^2)$, whose diagram were similar to leaf curves in a interval. On the basis ofgeometric meaning of derivative, $\cot(TA)$ can be interpreted as $f'(0)$ (value of leaf curvilinear equation'sderived function at origin of coordinates, thus, biological significance of leaf curvilinear equationparameters are showd in Table 2.

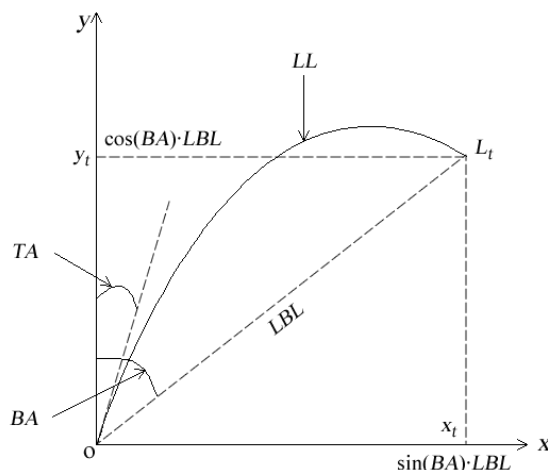
Table 2The various functions of leaf curve, their derivative function and the derivative value at the origin, and the biological significance of the parameters

$f(x)$	$f'(x)$	$f'(0)$	Biological significance of the parameters
ax^2+bx	$2ax+b$	b	$-a$:curvature; $b=\cot(TA)$
$ax^4+bx^3+cx^2+dx$	$4ax^3+3bx^2+2cx+d$	d	$d=\cot(TA)$
$asin(bx)$	$ab\cos(bx)$	ab	leaf curve peak: $(\pi/2b,a)$; $ab=\cot(TA)$
axb^x	$ab^x\log(b)x+ab^x$	a	$a= \cot(TA)$; $-b$:curvature
$(a+bx^c)/(d+x^c)$	$\frac{bcx^{c-1}}{x^c+d} - \frac{cx^{c-1}(bx^c+a)}{(x^c+d)^2}$	0	N/A
$(a+bx)/(1+cx+dx^2)$	$\frac{b}{dx^2+cx+1} - \frac{(bx+a)(2dx+c)}{(dx^2+cx+1)^2}$	$b-ac$	$b-ac=\cot(TA)$

From Table 2, wesawthat only quadratic function, sine function, and Hoerl-like function had specific biological significance for all the parameters: for quadratic function ax^2+bx , $-a$ expresses the leaf

curvature, and b expresses the cotangent value of leaf tangential angle; for sine function $a \sin(bx)$, a expresses the ordinate value of leaf curve peak point, $\pi/2b$ abscissa value of expresses the leaf curve peak point, and a/b expresses the cotangent value of leaf tangential angle; for Hoerl-like function axb^x , a expresses the cotangent value of leaf tangential angle, and $-b$ expresses the leaf curvature. So that we addressed them and validated the assumptions.

3.1.2.2 Solution of the Leaf Curvilinear Equation



L_t is leaf tip, x_t is the abscissa of L_t , y_t is the ordinate of L_t .

Fig. 3. The geometrical properties of leaf curve

As shown in Fig. 3, the leaf tip could be expressed as $(\sin(BA) \cdot LBL, \cos(BA) \cdot LBL)$ by solving $\triangle OL_t$. We substituted coordinates of origin and L_t into curvilinear function for curving leaves to get equation set as Eq. 3.

$$\begin{cases} f(\sin(BA) \cdot LBL) = \cos(BA) \cdot LBL \\ f'(0) = \cot(TA) \end{cases} \quad (3)$$

The equation sets and their solutions corresponding to the three curvilinear functions for curving leaves we supposed above were shown in Table 3.

Table 3 Three curvilinear functions for curving leaves and the corresponding equation sets and their solutions

$f(x)$	Equation set	Solution
ax^2+bx	$\begin{cases} f(\sin(BA) \cdot LBL) = a \cdot \sin(BA)^2 \cdot LBL^2 + b \cdot LBL \cdot \sin(BA) = \cos(BA) \cdot LBL \\ f'(0) = b = \cot(TA) \end{cases}$	$a = -\frac{\sin(BA) \cdot \cot(TA) - \cos(BA)}{LBL \cdot \sin(BA)^2}$ $b = \cot(TA)$
$a \sin(bx)$	$\begin{cases} f(\sin(BA) \cdot LBL) = a \cdot \sin(b \cdot LBL \cdot \sin(BA)) = \cos(BA) \cdot LBL \\ f'(0) = a \cdot b = \cot(TA) \end{cases}$	$a = \frac{LBL \cdot \cos(BA)}{\sin(b \cdot LBL \cdot \sin(BA))}$ $a \cdot b = \cot(TA)$
axb^x	$\begin{cases} f(\sin(BA) \cdot LBL) = a \cdot LBL \cdot \sin(BA) \cdot b^{LBL \cdot \sin(BA)} = \cos(BA) \cdot LBL \\ f'(0) = a = \cot(TA) \end{cases}$	$a = \cot(TA)$ $b = \left(\frac{\cos(BA)}{\sin(\sin(BA) \cdot \sin(TA))} \right)^{\frac{1}{\sin(BA) \cdot LBL}}$

As shown in Table. 3, quadratic function, and Hoerl-like function could be solved directly, but there was no analytical solution for sine function, and we wrote a C# program to calculate the approximate solution with a step of 10^{-5} cm.

3.1.2.3 Validation for Leaf Curvilinear Functions

We used the observed value of tangential angle, bowstring angle, and bowstring length for solving equation set, except leaf length. Meanwhile, the leaf length also could be calculated as the arc length between leaf basal and leaf tip by the formula of arc length. Therefore, we can validate the leaf curvilinear function through comparing the observed leaf length with the calculated arc length. The three leaf curvilinear functions were validated by 1:1 line of observed and calculated leaf length (Fig. 4), and the statistical parameters were shown in Table 4.

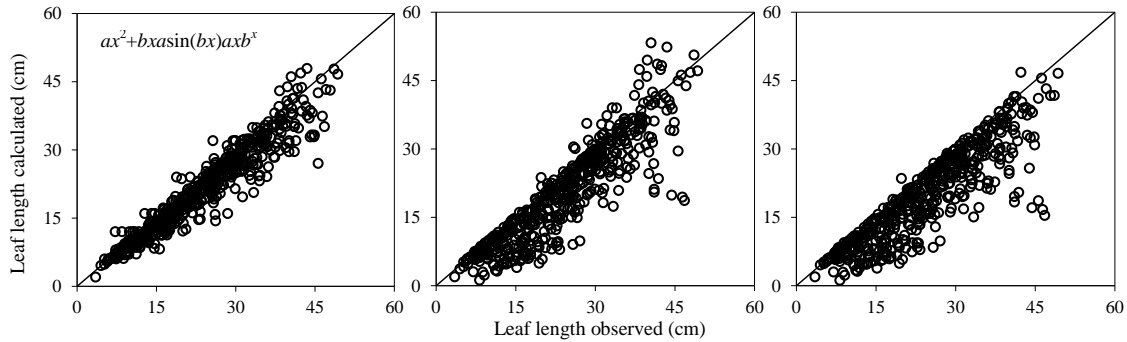


Fig. 4. Comparison of the observed and the stimulated leaf length on main stem in 2011-2012

Table 4 Comparison of statistical parameters of the observed and the stimulated leaf blade length on main stem in 2011-2012

$f(x)$	Statistic parameters of simulation and observation					
	n	$d_a(\text{cm})$	$d_{ap}(\%)$	$RMSE(\text{cm})$	r	Sig.
ax^2+bx	509	2.521	10.426	3.974	0.956 ^{***}	$r_{(507,0.001)}=0.145$
$asin(bx)$	509	3.730	15.427	6.083	0.904 ^{***}	$r_{(507,0.001)}=0.145$
axb^x	509	4.693	19.410	6.749	0.889 ^{***}	$r_{(507,0.001)}=0.145$

The results showed that the best fitted one (with the highest r and lowest d_a , d_{ap} , and $RMSE$, Table 4) with specific biological significance (Table 2) for leaf curve was quadratic function, so that the curvilinear model for curving leaves could be described as a quadratic function as Eq. (4).

$$f(x) = -Lc_i \cdot x^2 + \cot(TA) \cdot x \quad (4)$$

where, Lc_i is the leaf curvature on the i th day after emergence.

3.1.2.4 Biomass-Based Leaf Curvature Simulation

The data in the experiments showed that quadratic coefficients of leaf curvilinear equations increased with the increase of corresponding leaf dry weight from leaf fully expanded until senescence. It means that leaf curvature decreased with the corresponding leaf biomass (Fig. 5), and could be fitted by reciprocal function with significant r (V1: $r=0.974$, $P<0.001$, $n=34$, $r_{(32,0.001)}=0.539$; V2: $r=0.637$, $P<0.001$, $n=51$, $r_{(49,0.001)}=0.447$. Table 5).

$$Lc_i = \frac{-Lc_a}{DWLB_i} - Lc_b \quad (5)$$

where $DWLB_i$ is the dry weight on the i th day after emergence; Lc_a and Lc_b are model parameters whose values and testing data shown in Table 5.

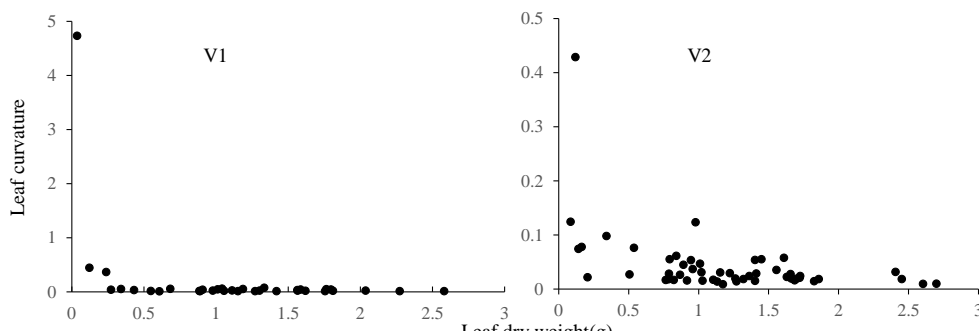


Fig. 5. Changes in the leaf curvature with the leaf dry weight in 2011-2012

Table 5. Significance test of the leaf curvature model and its parameter

Cultivar	<i>n</i>	<i>r</i>	Sig. for <i>r</i>	Sig. for <i>F</i>	Parameter symbolic	Unstandardized coefficients	<i>t</i>
V1	34	0.974***	$r_{(32, 0.001)}=0.539$	0.000	Lc_a	-0.165	-24.127***
					Lc_b	0.163	4.625***
V2	51	0.637***	$r_{(49, 0.001)}=0.447$	0.000	Lc_a	-.018	-5.789***
					Lc_b	-0.015	-1.826

3.2 Validation

The models developed above were validated with the independent datasets from Exp. 2 and Exp. 3, and the results showed that the correlation (*r*) of simulation and observation probability for straight leaves and curvature for curving leaves all had significant level at $P < 0.001$, and that the average absolute difference (d_a), the ratio of d_a to the average observation (d_{ap}), and the root mean square error (*RMSE*) all were smaller (Table 6). Fig. 6 and Fig. 7 indicated that the observed and simulated probability for straight leaves and curvature for curving leaves were all close to the 1:1 line.

Table 6 Comparison of statistical parameters of simulation and observation in the probability for straight leaves and curvature for curving leaves on main stem in 2012-2013

Models	Cultivar	Statistic parameters of simulation and observation					
		<i>n</i>	d_a	$d_{ap}(\%)$	<i>RMSE</i>	<i>r</i>	Sig.
Probability for straight leaves	V1, V2	52	0.001	0.245	0.191	0.762***	$r_{(50, 0.001)}=0.443$
Curvature for curving leaves	V1	93	0.007	-9.196	0.060	0.648***	$r_{(91, 0.001)}=0.336$
	V2	99	-0.005	-7.000	0.042	0.541***	$r_{(97, 0.001)}=0.326$

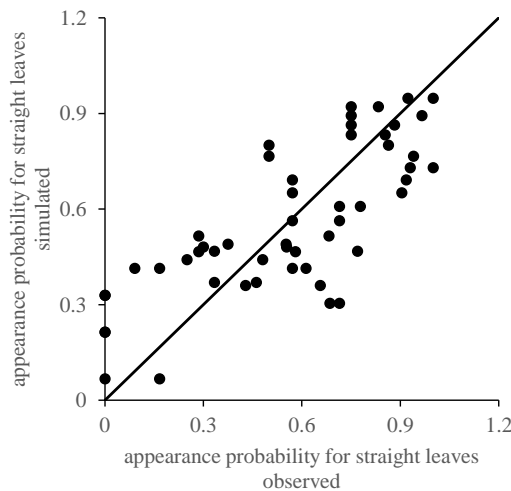


Fig. 6. Comparison of the observed and the stimulated probability for straight leaves in 2012-2013

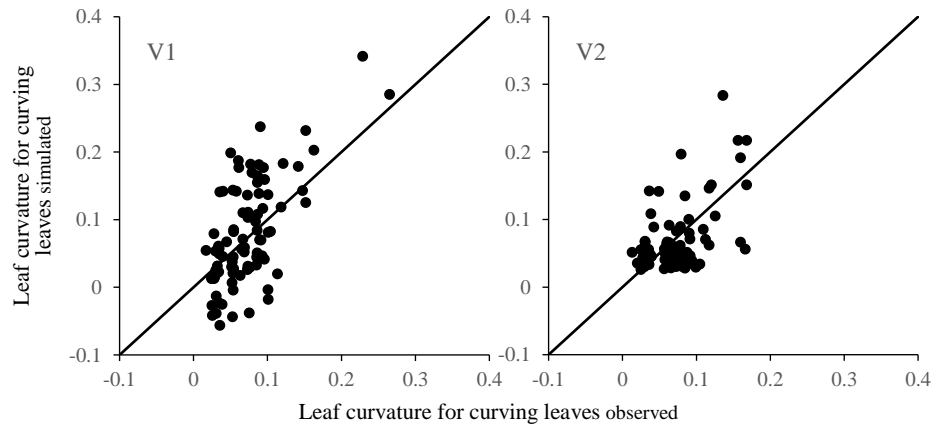


Fig. 7. Comparison of the observed and the stimulated leaf curvature for curving leaves in 2012-2013

4 Discussion

4.1 Establishment of Biomass-Based Leaf Curvilinear Model Will Help to Develop FSRM

Functional-structural plant model with better mechanistic embodies the interaction between plant morphogenesis and cultivars and environmental factors by integrated the functions of growth model with the structures of morphological model [26]. So far, there were a lot of research on modeling the leaf curve [10-21] which aimed at 3D reconstruction of crop canopy with a high precision. They well explained the effects of structures on functions by combining technical instrumentalities like geometrical ray trace [27, 28], but could not change the structures as a response of the changed functions. Shi et al. [22] linked leaf curve with corresponding dry weight through force analyzing on rice leaf, but the leaf curvilinear equation which was limited by a form of second order differential equation in this research was difficult to use. Groer et al. [29] established a dynamic 3D model of rapeseed using the modelling language XL [30] and made the morphological model response of different nitrogen levels using a sinks and source system like GREENLAB and the LEAFC3-N model [31] for photosynthesis at different N-regimes. But the time from sowing to the rosette stage and the conditions except nitrogen was neglected in this model. Jullien et al. [32] constructed a FSPM by characterizing the interactions between architecture and source-sink relationships in winter oilseed rape using the GREENLAB model. However, it mainly considered the relationship between biomass and leaf area [33], and the description of blade shape was relatively simple. Cao et al. [34] and Zhang et al. [25] established models to meticulously predict leaf morphological parameters like leaf length, width, and angles, apart from leaf curve. We developed biomass-based leaf curvilinear model for rapeseed by linking the leaf curvature with the corresponding leaf biomass, which could realize combination of structures with functions, explained effects of environmental conditions on leaf morphogenesis, and set the stage for the development of functional-structural rapeseed models.

4.2 The Research Provided a Mechanistic and Universal Method for Leaf Curve Modeling

As to facilitate observation and simulation, we set the leaf into a Cartesian coordinate system with the leaf basal point as the origin and growth direction of main stem as y-axis. To make the model more precisely, two cases of curving characteristic (straight or curving) were analyzed. In order to determine the form

of leaf curvilinear equation to be difficult to measure directly, we compared various functions based on their biological significance, and calculated and observed leaf length. For eliminating the apparent differences between treatments and replications, we normalized the leaf rank into the interval of (0, 1]. All these practices made the model to have better mechanistic and universal.

4.3 Models Developed In This Paper Needs To Be Improved

The biomass-based rapeseed leaf curvilinear model developed in this paper for the stage from leaf fully expanded until senescence, but the processes of leaf extension, senescence, and distorting is neglected. Thus, it needs to be studied further.

5 Conclusions

This paper presents a biomass-based leaf curvilinear model for rapeseed designed to explain effects of cultivars and environmental conditions on leaf curve. Various model variables, including leaf biomass, length, and angles were parameterized based on datasets derived from the experiments with rapeseed cv. Ningyou 18, and Ningza 19.

With the help of our descriptive model, it will be easy to fulfill calculation of leaf curve via biomass, canopy structure via leaf curve, light distribution and the photosynthesis via canopy structure, and biomass via photosynthesis. It should be possible to connect morphological model with physiological model via biomass, and to develop the FSRM.

A similar method could also be applied in other crops like maize and rice, and it could help to the regulation and selection for ideal plant type in the future.

Acknowledgments

This work is supported by the grants from the National Natural Science Foundation of China (31171455, 31201127, and 31471415), the National High-Tech Research and Development Program of China (2013AA102305-1), the Jiangsu Agriculture Science and Technology Innovation Fund of China [CX(12)5060], and Jiangsu Academy of Agricultural Sciences Basic Scientific Research Work Special Fund [ZX(15)2008, ZX(15)4001].

References

1. Cao Hongxin, Zhang Chunlei, Li Guangming, et al. Researches of optimum shoot and ramification number dynamic models for rapeseed (*Brassica napus* L.)[C]. World Automation Congress (WAC), 2010, pp.129 - 135. IEEE.
2. FAO Statistics Division. FAOSTAT [DB/OL]. <http://faostat.fao.org/>, 2015.
3. Cao Hongxin, Zhang Chunlei, Li Guangming, et al. Researches of optimum leaf area index dynamic models for rape (*Brassica napus* L.)[J]. Computer & Computing Technologies in Agriculture II, 2009, 3, 1585.
4. Hama S, Kondo A. Enzymatic biodiesel production: An overview of potential feedstocks and process development[J]. Bioresource Technology, 2013, 135(2):386–395.
5. Kropff M J, Laar H H V, Matthews R B. ORYZA1: An ecophysiological model for irrigated rice production[C]. Sarp Research Proceedings Pp, 1994, (1994).
6. Boote KJ, Pickering NB. Modeling photosynthesis of row crop canopies[J]. HortScience,

- 1994, 29(12): 1423-1434.
7. Gao Liangzhi, JinZhiqing, ZhengGuoqing, et al. Wheatcultivational simulation-optimization-decision marking system (WCSODS) [J]. Jiangsu Journal of Agricultural Sciences, 2000, 16(02): 65-72. (in Chinese)
8. Tang Liang, Zhu Yan, HannawayD, et al. RiceGrow: A rice growth and productivity model [J]. NJAS - Wageningen Journal of Life Sciences, 2009, 57(1): 83-92.
9. Wang W, Li Z, Su H. Comparison of leaf angle distribution functions: Effects on extinction coefficient and fraction of sunlit foliage [J]. Agricultural and Forest Meteorology, 2007, 143(1-2): 106-122.
10. Stewart D W, Dwyer L M. Mathematical characterization of maize canopies [J]. Agricultural and Forest Meteorology, 1993, 66(3-4): 247-265.
11. GuoYan, LiBaoguo. Mathematical description and three-dimensional reconstruction of maize canopy [J]. Chinese Journal of Applied Ecology, 1999, 10(01): 39-41. (in Chinese)
12. Zhao Chunjiang, Zheng Wengang, GuoXinyu, et al. The computer simulation of maize leaf [J]. Journal of Biomathematics, 2004, 19(04): 493-496. (in Chinese)
13. Zheng Wengang, GuoXinyu, Zhao Chunjiang, et al. Geometry modeling of maize leaf canopy [J]. Transactions of the CSAE, 2004, 20(01): 152-154. (in Chinese)
14. Liu Hongwei, Wu Bin, Zhang Hongying, et al. Research on rice leaf ceometric model and its visualization [J]. Computer Engineering, 2009, 35(23): 263-264+268. (in Chinese)
15. Yang Hongyun, Luo Wei, He Huojiao, et al. 3D Morphology modeling and computer simulation of rice main stem [J]. ActaAgriculturaeUniversitisJiangxiensis, 2008, 30(06): 1153-1156+1160. (in Chinese)
16. LuoWei, Yang Hongyun, Deng Song, et al. A study based on multi-segment & proportion geometrical model of rice leaf and 3D morphology modeling [J]. ActaAgriculturaeUniversitisJiangxiensis, 2009, 31(05): 970-974. (in Chinese)
17. ChenGuoqing. Studies on simulation and visualization of morphogenesis in wheat [D]. Nanjing Agricultural University, 2004. (in Chinese)
18. DornbuschTino, WerneckePeter, DiepenbrockWulf. Description and visualization of graminaceous plants with an organ-based 3D architectural model, exemplified for spring barley (*Hordeumvulgare* L.) [J]. The Visual Computer, 2007, 23(8): 569-581.
19. Zhang Yonghui, Tang Liang, Liu Xiaojun, et al. Dynamic simulation of leaf curve in rice based on Gaussian function [J]. ScientiaAgricolturaSinica, 2013, 46(01): 215-224. (in Chinese)
20. EspañaMaría Luisa, BaretFrédéric, Aries Franck, et al. Modeling maize canopy 3D architecture: Application to reflectance simulation [J]. Ecological Modelling, 1999, 122(1-2): 25-43.
21. Watanabe Tomonari, HananJim S, Room Peter M, et al. Rice morphogenesis and plant architecture: measurement, specification and the reconstruction of structural development by 3D architectural modelling [J]. Annals of Botany, 2005, 95(7): 1131-1143.
22. Shi Chunlin, Zhu Yan, Cao Weixing. A quantitative analysis on leaf curvature characteristics in rice [J]. ActaAgronomicaSinica, 2006, 32(05): 656-660. (in Chinese)
23. ZhengBangyou, Shi Lijuan, Ma Yuntao, et al. Three-dimensional digitization in situ of rice canopies and virtual stratified-clipping method [J]. ScientiaAgricolturaSinica, 2009, 42(04): 1181-1189. (in Chinese)

24. Cao Hongxin, HananJim Scott, Liu Yan, et al. Comparison of Crop Model Validation Methods [J]. *Journal of Integrative Agriculture*, 2012, 11(8): 1274-1285.
25. Zhang Weixin, Cao Hongxin, Zhu Yan, et al. Morphological Structure Model of Leaf Space Based on Biomass at Pre-Overwintering Stage in Rapeseed (*Brassica napus* L.) Plant [J]. *Acta Agronomica Sinica*, 2015, 41 (02): 318-328. (in Chinese)
26. Cao Hongxin, Zhao Suolao, Ge Daokuo, et al. Discussion on development of crop models [J]. *Sci Agric Sin*, 2011, 44: 3520–3528 (in Chinese)
27. Chelle Michaël, Saint-Jean Sébastien. Taking into account the 3D canopy structure to study the physical environment of plants: the montecarlo solution [C]. 4th International Workshop on Functional-Structural Plant Models, 2004: 176-180.
28. Ma Yuntao, Wen Meiping, Li Baoguo, et al. Efficient model for computing the distribution of direct solar radiation in maize canopy at organ level [J]. *Transactions of the CSAE*, 23(10): 151-155. (in Chinese)
29. Groer Christian, Kniemeyer Ole, Hemmerling Reinhard, et al. A dynamic 3D model of rape (*Brassica napus* L.) computing yield components under variable nitrogen fertilization regimes [C]. 5th International Workshop on Functional-Structural Plant Models, November 2007: 4.1-4.3.
30. Kniemeyer O. Rule-based modelling with the XL/GroIMP software [C]. *Proceedings of 6th GWAL*, April 14-16, 2004. pp 56–65.
31. Müller J, Wernecke P, Diepenbrock W. LEAFC3-N: a nitrogen-sensitive extension of the CO₂ and H₂O gas exchange model LEAFC3 parameterised and tested for winter wheat (*Triticum aestivum* L.) [J]. *Ecol Model*, 2005, 183: 183–210.
32. Jullien Alexandra, Mathieu Amélie, Allirand Jean-Michel, et al. Characterization of the interactions between architecture and source-sink relationships in winter oilseed rape (*Brassica napus*) using the GreenLab model [J]. *Ann Bot*, 2011, 107(5): 765-779.
33. Jullien Alexandra, Allirand Jean-Michel, Mathieu Amélie, et al. Variations in leaf mass per area according to N nutrition, plant age, and leaf position reflect ontogenetic plasticity in winter oilseed rape (*Brassica napus* L.) [J]. *Field Crops Research*, 2009, 114(2): 188-197.
34. Cao Hongxin, Zhang Wenyu, Zhang Weixin et al. Biomass-based rapeseed (*Brassica napus* L.) leaf geometric parameter model [C]. *Proceedings of the 7th International Conference on Functional-Structural Plant Models*, 9 - 14 June, 2013: 26.